

Review

Fermented Foods Strengthen Immunity against Viral Infections

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Received: 28 April 2025; Revised: 21 May 2025; Accepted: 12 June 2025; Published: 12 August 2025

Abstract: Fermented foods have been shown to exert positive effects on gut health and immune function. However, the potential of fermented foods to enhance the bioavailability of bioactive compounds and support the growth of the beneficial microbial community's key factors in antiviral immunity remains less explored. In this review, we show that probiotic-fermented food improves the bioactive compound contents and is increasingly studied by basic and clinical researchers. Bioactive compounds, including phenolic, alkaloids, terpenoids, flavonoids, stilbenes, coumarins, tannins, anthocyanidins, flavones, isoflavonoids, and polyphenols, are increased in the probiotic fermentation conditions. Additionally, beneficial bacteria such as *Lactobacilli*, *Bifidobacteria*, *Pediococcus*, and *Weissella* are also restored in the fermented foods. These bioactive compounds, combined with a functional microbiota, play a role in preventing viral infections by targeting influenza, noroviruses (NoVs), Murine norovirus-1 (MNV-1), and COVID-19, while also stimulating the immune function of the host. It was suggested that clinical and pre-clinical investigations are required to explore the dose-response and duration efficacy of probiotic fermented foods against viral infections.

Keywords: fermented food; probiotics; bioactive compounds and antiviral potential

1. Introduction

Fermented foods have been produced and utilized as a diet since the development of human civilizations [1]. It is valuable for human health by relieving the blood cholesterol levels, and providing protection against pathogens, hazardous, and carcinogenic substances. Additionally, improving the digestibility for individuals with lactose intolerance (the fermentation process breaks down lactose into simpler sugars) [2,3]. The health-promoting effects of fermented foods are largely attributed to their rich content of bioactive compounds and beneficial native microbiota. These components act synergistically to enhance the nutritional profile and support overall human health and well-being [4]. For example, polyphenols, flavonoids, and antioxidants possess various healthpromoting properties, including anti-inflammatory, anti-cancer, and immune-boosting effects [5–7]. Furthermore, angiotensin-1-converting enzymes [8], valyl-prolyl proline, and isoleucyl-prolyl-proline inhibitors have also been observed in fermented food [9]. These compounds can potentially treat hypertension and other associated health disorders. Similarly, the bioactive compounds and probiotic strains were found in the fermented food with antiviral potential and various mechanisms of action [3]. The application of these bioactive compounds and probiotics has shown promising efficacy against a wide range of viral infections, as demonstrated by both in vitro and in vivo studies [10,11]. These agents can enhance the host immune response through direct antiviral activity or by modulating immune function. Microbial fermentation provides a cost-effective and sustainable method for producing such beneficial metabolites, as it bypasses the need for complex pre-purification or hydrolysis steps required by other production techniques [12]. The fermentation can generate diverse amino acids; acidic, basic, or hydrophobic, that contribute not only to the nutritional profile but also to the aroma and functionality of the bioactive compounds in fermented foods [4,7,13]. Additionally, studies have shown that whey [14], and yogurt [15] fermented with Lactobacillus yield bioactive compounds that inhibit ACE (angiotensin-converting enzyme) activity, thereby modulating the function of toll-like receptor-4. Furthermore, research has identified a bioactive peptide (P18) produced by Bacillus subtilis within legume-based fermented foods [16]. This constitutes a



significant component of the microbiota. Microflora of the Bifidobacteria and Lactobacilli might have the capability as anti-viral. For example, cell-free supernatants of Lactobacillus spp., Streptococcus thermophilus, and Bifidobacterium bifidum cultures in yogurt produce metabolites that can prevent influenza virus infection [17,18]. Moreover, Lactobacillus delbrueckii subsp. bulgaricus releases the bacteriocin and was also found as an antiinfluenza [18,19]. Clinical trials showed that the mucosal cells (IL-6 and IL-10) increase were observed in the clinical trials by using fermented food rich in bioactive and probiotics [20]. During the COVID-19 outbreak, several reports suggested that lower mortality rates observed in regions such as Asia, Africa, and parts of Europe may be partially linked to the widespread consumption of fermented foods in these populations [13,21–23]. In this review, we have thoroughly examined the bioavailability of bioactive compounds and the composition of probiotic bacterial communities in various fermented foods, highlighting their potential health-promoting benefits. Specifically, we analyzed how probiotic-fermented foods strengthen the body's defenses against viral infections, including respiratory ailments, gastrointestinal tract disorders, as well as infections caused by herpes simplex viruses. Notably, we have also examined emerging research regarding the potential role of fermented foods, including their probiotic components, in mitigating the impact of COVID-19. Moreover, this review provides a thorough analysis of the safety aspects surrounding fermented foods, addressing concerns related to contamination and spoilage, and outlining strategies for maintaining their integrity and nutritional value throughout the fermentation process and storage.

2. Microbiome and Functional Metabolites of Fermented Foods

Food fermentation and preservation methods significantly influence the flavor, texture, and shelf life of food products through various chemical transformations. Fermented foods derived from plant-based sources, dairy, alcoholic beverages, and marine products are widely consumed across the globe. Among these, fermented vegetables and fruits hold a prominent place in the history of human civilization, reflecting a long-standing tradition of dietary and cultural significance [1]. For example, sauerkraut (fermented cabbage), was commonly consumed in the Roman Empire [24], while *Jiangshui*, a traditional fermented vegetable in China, and Kimchi in Korea have been integral components of regional diets for centuries [25,26]. The fermented soybeans called "Miso and Natto" are used in Japan, while Indonesian Tempeh" [27] has different impacts on human health. Similarly, fermented alcoholic beverages from fruits, cereals, and milk were prepared under culture-dependent and independent techniques. For example, the fermentation of sake, flavored liquor, grape wine, and alcohol [28–30] under different cultures and parameters. Fermented food is a rich source for metagenomics, metaproteomics, metabolomics, and diverse meta-analyses studies, offering a plethora of opportunities for in-depth exploration. The various analyses employed to investigate fermented foods include:

2.1. Functional Microbiota

Dairy-fermented products have increased attention and are integral components of diets across the globe, as recognized by the International Dairy Federation (IDF) [31]. These products have been associated with potential health benefits, including enhanced insulin sensitivity, reduced cholesterol levels, and improved blood pressure regulation. Airag, a traditional fermented beverage from Mongolia made from unpasteurized mare's milk, has long been used for its purported therapeutic properties [32]. Probiotic strains such as Streptococcus thermophilus and Lactobacillus delbrueckii, commonly found in yogurt, contribute to improved digestion and the restoration of gut microbiota balance [29]. Overall, fermented foods are rich in probiotic bacterial strains and are utilized for various health-promoting functions, as outlined in Table 1. The microbial diversity in the fermented foods investigated, such as Firmicutes and Proteobacteria, were the pre-dominant phyla in the cheese, jueke, and koumiss. Similarly, Lactobacillus, Leuconostoc, Weissella, Enterococcus, and Pediococcus were observed in fermented vegetables and fruits [33,34]. In the cheese and Kefir samples, collectively Lactococcus, Lactobacillus, Streptococcus, Acetobacter, and Leuconostoc bacterial strains were found dominant [35]. Wu et al. demonstrated that L. casei, L. helveticus, and Lactobacillus plantarum were increased in koumiss [36]. Furthermore, Enterococcus faecalis, Lactococcus lactis, Leuconostoc mesenteroides, Lactobacillus plantarum, L. casei, and L. zeae were reported in the Chinese sauerkraut [37]. It is suggested that the isolated probiotic strains from fermented food could be employed for basic and clinical research. Additionally, there is increasing focus on the demand for probiotics in the financial industry [1], owing to their physiological functions and potential to treat to different disorders. Yogurt, nutrition bars, snacks, infant foods, and numerous other products are now being fermented with probiotic bacterial strains to enhance their nutritional contents. For, example, gastroenterologists often prescribe commercial lyophilized pills for their patients. However, the scientific validation of probiotics in fermented foods for improving human health functions remains largely unexplored.

Table 1. Fermented foods and their impact on human health and host spots for various probiotic strains.

| Fermented Food | | | Functions | References |
|-------------------------------|--|--------------------------------|---|------------|
| Koumiss | Lactobacillus coryniformis, L. paracasei, L. kefiranofaciens L. curvatus L. fermentum, L. casei, L. helveticus, Lactobacillus plantarum | 11. | Resistance to the low-acidic level Antimicrobial activities | [36] |
| Kimchi | L. casei DK128, Lactobacillus buchneri UA-1 and UA-2 | i. ii. iii. | IgG antibodies rapid initiation. Induce innate immune cells and cytokines Metabolized uric acid | [38,39] |
| Jiangshui | Lactobacillus, Limosilactobacillus fermentum, and L. bacilli | i. ii. | Effective in hyperuricemia and gout Provides coldness to the body | [25] |
| Yogurt | Bifidobacterium animalis subsp. lactis | i. ii. iii. iv. v. | Suppressing infection of helicobacter pylori Lower the cholesterol levels in the serum Stimulate the immune function Improve lactose metabolism Antimicrobial, anticarcinogenic, antimutagenic, and antidiarrheal properties | [40,41] |
| Kefir | L. acidophilus, Lactobacillus delbrueckii subsp. bulgaricus, Streptococcus thermophilus, L. crispatus, L. gasseri, L. jensenii, L. rhamnosus | | Act as lactose-tolerant and antibacterial Antidiabetic Suppressing tumors, and acting as an anti- hypertensive, anti-inflammatory, antioxidant, and carcinogenic | [40] |
| Mango pickle, Naan | Indigenous microflora, yeast | i. ii. | Appetizer and meal purposes Increases elasticity in blood vessels and produce new blood cells | [42] |
| Sourdough (Khamir) | Enterococcus mundtii, and Wickerhamomyces anomalus, Bacillus subtilis LZU-GM | i. ii. iii. | Contains gluten-degrading probiotics Lowers the risk of cancer, aging, and arthritis Relieves celiac disease | [43,44] |
| Fermented milk, | Acinetobacter, Enterobacteriaceae, and Aeromonadaceae | i. ii. | Immunomodulation and amelioration of colitis and diabetes Used for gastrointestinal disorders, cancer, high cholesterol, and blood pressure | [45] |
| Shubat and Ayran | Leuconostoc and Enterococcusgenera | _ | gulates bile salt tolerance and increases bodies susceptibility | [46] |
| PaoCai | Enterococcus faecalis, Lactococcus lactis, Leuconostoc mesenteroides, L. plantarum, L. casei and L. zeae | | Appetizers and vitamin absorption Anticancer effects | [37] |
| Fermented Portuguese olive | L. plantarum and L. paraplantarum | i. ii. iii. | Affects acid and bile salt-tolerance Simulated digestion Exopolysaccharide producing abilities | [47,48] |
| Koumiss | L. helveticus and L. delbrueckii | i. ii. | Treat urogenital tract diseases and inflammatory disorders Source of the biogenic amines | [49] |
| Raw camel milk | L. fermentum, L. plantarum, L. casei, Lactococcus lactis, Enterococcus faecium, and S. thermophilus | i. ii. | Monitoring diabetes including cholesterol levels, liver and kidney ailment Oxidative stress decreases and heals the wound. | [50] |
| Pickle and cucumber | | i. ii. | Prolongs shelf life Reduces the toxic levels of protein and minerals | [42] |
| Tarhana | Streptococcus thermophilus, L. fermentum, Enterococcus faecium, Pediococcus | i. ii. | Rich source of minerals and nutrients Affect pathogens and enhances shelf life | [51] |

| | pentosaceus, Leuconostoc pseudomesenteroides, Weissella cibaria, L. plantarum, L.delbrueckii, Leuconostoc citreum, L. paraplantarum and L casei. | | | |
|--|---|-------------------|--|---------|
| Meju, Doenjang, Jeotgal, and Mekgeolli | L.mesenteroides, L.plantarum, Aspergillus, Bacillus, Halomona sp., Kocuria sp., and Saccharomyces cerevisiae | i. s 11. | Anticancer, anti-obesity, Antioxidant, anti-inflammatory, and anti- diabetes | [52] |
| Dhokla | L. plantarum and Weissella cibaria | i. ii. iii. | Antimicrobial activity Infuence bile tolerance Antibiotic susceptible | [53] |
| Cheese | L. lactis L. delbrueckiiL. helveticus, L. casei, L. plantarum, L. salivarius, Leuconostoc spp., S. thermophilus, Ent. durans, Ent. faecium, Staphylococcus Brevibacterium linens, Propionibacterium freudenreichii, Penicillium camemberti, P. roqueforti | i. ii. iii. | Contains-aminobutyric acid (GABA). Exopolysaccharides, peptides, vitamins, fatty acids, and organic acids Anticancer effects | [54–56] |
| Pla-khao-sug | Ped. cerevisiae, L. brevis, Staphylococcus sp., Bacillus sp. | i. ii. | Boost the immune system Suppression of pathogenic bacteria | [57] |
| Tapai Ubi | Saccharomycopsis fibuligera, Amylomyces rouxii, Mu. circinelloides, Mu. javanicus, Hansenula spp, Rhi. arrhizus, Rhi. oryzae, Rhi. Chinensis | i. ii. | Useful for nerve and muscle cells Balancing microbial diversity and improving immunity | [58,59] |
| Tungrymbai | B. subtilis, B. licheniformis, B. pumilus | i. ii. | Isoflavone transformation and high antioxidants Cheap source of proteins | [60,61] |
| Thua nao | B. subtilis, B. pumilus, Lactobacillus sp. | i. ii. | Contains pyrazine compounds and is rich in nutrients Proteins supplement | [62,63] |
| Sufu | Actinomucor elenans, Mucor. silvatixus, Mu. corticolus, Mu. hiemalis, Mu. praini, Mu. racemosus, Mu. subtilissimus, Rhiz. Chinensis | | Considered a healthy food because of its low cholesterol content Act as sources of protein and calcium | [64,65] |
| Miso | Ped. acidilactici, Leuc. paramesenteroides, Micrococcus halobius, Ped. halophilus, Streptococcus sp., Sacch. rouxii, Zygosaccharomyces rouxii, Asp. Oryzae | ii. | Reduces cancer cell growth Improves the digestive system Lowers cholesterol levels. | [24,66] |
| Koozh and gherkin | Lactobacillus and Weissella | i. ii. | Achieves the maximum cholesterol reduction. Exopolysaccharide source | [34] |
| Airag | L. helveticus, L. kefiranofaciens, Bifidobacterium mongoliense, and Kluyveromyces marxianu | i. | Reduces thirst and hunger Improves metabolism Treats heart and lung diseases | [32] |
| Chhurpi | L. farciminis, L. paracasei, L. biofermentans, L. plantarum, L. curvatus, L. fermentum, L. alimentarius, L. hilgardii, W. confusa, Ent. faecium, Leuc. Mesenteroides | Coı | nstains high protein content and carbohydrates low fat levels | [67,68] |
| Somar | L. paracasei, L. Lactis | | | [69] |

| Boza | Lactobacillus sp., Lactococcus sp., Pediococcus sp., and Leuconostoc sp., | Contains biogenic amine content | [70,71] |
|---------------------------|---|---|---------|
| Suan-tsai and fu- tsai | Ent. faecalis, L. alimentarius, L. brevis, L. coryniformis, L. farciminis, L. plantarum, L. versmoldensis, Leuc. citreum, | | [72] |
| Nem-chua | L. pentosus, L. plantarum, L. brevis, L. paracasei, L. fermentum, L. acidipiscis, L. farciminis, L. rossiae, L. fuchuensis, L. namurensis, Lc. lactis, Leuc. citreum, Leuc. fallax, P. acidilactici, P. pentosaceus, P. stilesii, Weissella cibaria, W. paramesenteroides | Inhibit the entrance of pathogenic microbes | [73] |

2.2. Detection of Bioactive Compounds in Fermented Food

Analytical instruments, including gas chromatography with mass spectroscopy (GC-MS), High-performance liquid chromatography (HPLC), High-Resolution Nuclear Magnetic Resonance (H-NMR), and ultra-highperformance liquid chromatography quadrupole time-of-flight mass spectrometry (UHPLC-Q-TOF MS/MS, have been used mainly for the food metabolites analysis (Table 2). The bioactive compounds, including isothiocyanates and hexanoic acid, were found in the nozawana zuke in higher concentrations, while acetic acid, acetoin, and 2,3butanedione were observed in low levels [74]. Barley fermented with L. plantarum dy-1 increased the indole-3lactic acid, phenyl lactic acid, homovanillic acid, and cafestol, in contrast, amino acids, nucleotides, saccharides, and other organic acids declined [9]. 2,4-di-tert-butylphenol, fatty acid esters, and sugar derivatives were reported by using the GC-MS analysis in the fermented whole grain. Similarly, in potherb mustard pickle, volatile compounds such as allyl, butenyl, isobutanol, and phenyl ethyl groups were identified as substituents in the side chain (R-groups) [75]. Furthemore, γ-aminobutyric acid (GABA), acetoin, acetoacetate, cellobiose, and alanine were detected in the fermented cantaloupe using H-NMR [12]. The integrated lipomics and metabonomics methods were employed in the fermented milk. The results showed that 108 metabolites and 174 lipids were reported. A total of 35 bioactive compounds were observed in the L. plantarum P9 additive fermented milk, among which the high levels of detected compounds were fatty acids, peptides, and celluloses [76]. The efficient and sustainable fermented foods were identified through multiple meta-omics tools, as shown in Table 2.

Table 2. Bioactive compounds from fermented foods by using various techniques.

| Fermented Foods | Metabolites | Techniques Use | References |
|---------------------------------|---|---------------------|------------|
| Fermented cantaloupe juice | Isoleucine, valine, lactic acid, alanine, β-alanine, sucrose, erythritol, gluconic acid, GABA, alpha-aminobutyric acid, methionine, acetoin, acetoacetate, and phenylpropanoic acid | H-NMR | [77] |
| Fermented soybeans | Glucosyringic acid, engeletin, dihydroxy-4-phenyl coumarin, histidine, leucine, lysine, methionine, phenylalanine, and tryptophan | UPLC- QTOF MS/MS | [75] |
| Nozawana-zuke | Isothiocyanates, hexanoic acid, lactic acid, acetic acid, acetoin, 2,3-butanedione, glutamine, valine, leucine, isoleucine, choline, and methionine | NMR, SPME- GC/MS | [74] |
| Fermented milk | Fatty acids, peptides, amino acids, carbohydrates, vitamins, aldehydes, ketones | UPLC-QTOF- MS/MS | [76] |
| Fermented coffee brews | aromatic amino acid, catabolites, and hydroxy dodecanoic acid | LC-Q-TOF-MS/MS | [78] |
| Fermented camel and bovine milk | Fatty acyls, benzenoids, organ heterocyclic, organic acids and derivatives, phenylpropanoids, polyketides, glycerophospholipids, sterol lipids, polyketides, prenol | UPLC-QTOF | [79] |

| | lipids, organic oxygen, glycerolipids, organooxygen, | | |
|----------------------|---|----------------|-------|
| | alkaloids and derivatives, sphingolipids, | | |
| | hydrocarbons, nucleosides, nucleotides, and lignans, | | |
| | neolignans and related compounds, organosulfur | | |
| | compounds, hydrocarbon derivatives, organic nitrogen | | |
| | compounds | | |
| | 1-stearoyl-lysophosphatidylcholine, gaboxadol, | | |
| | guanine, cytosine, 4-acetamido benzoic acid, | | |
| Farmantad agat mills | taurochenodeoxycholic acid, 2,6- | Q-HRMS-UPLC | F001 |
| Fermented goat milk | dimorpholinopyrimidine-4-carboxylic acid, D-proline, | Q-IIKMS-UFLC | [80] |
| | DL-Glutamic acid, O-beta-D-glucosyl-trans-zeatin, | | |
| | N2-1-Carboxyethyl-N5 diamino methylene ornithine, | | |
| | Citric acid, pipecolic acid, glutamic acid, Isoleucine, | | |
| Meju | Leucine, methionine, phenylalanine, tyrosine, proline, | UPLC-OTOF-MS | [81] |
| - j | threonine, valine | (| [] |
| | Volatiles (5 alcohols, dodecanoic acid, and 1,3- | | |
| Cereal-based | hexadiene) and the polyphenolic compounds gallic | | |
| fermented foods | acid, epigallocatechin-gallate, epigallocatechin, | SPME-GC | [82] |
| Termented foods | flavonoids, protocatechuic acid, and total polyphenols | | |
| | Amino acids, Organic acids, Sugars, Amines, Phenolic | | |
| Soymilk fermented | | H-NMR | [83] |
| | compounds, Lipids, Choline, and Trigonelline | | |
| | Galactosamine, Maltose, Phenylacetic acid, | | |
| | Cuminaldehyde, Adenosine, Glucose 1-phosphate, | | |
| Fermented Barley | Cafestol, Aspartic acid, Lysine, Tryptophan, Citric | UPLC-HRMS | [9] |
| , | acid, Glucose Asparagine, Docosahexaenoic acid | | L- 3 |
| | methyl ester, Histidine, Tyrosine, D-glucosamine-6- | | |
| | phosphate, Arginine, Fumaric acid, Benzaldehyde | | |
| Gochujangs | Amino acids, organic acids, sugar and sugar alcohol, | UPLC-QTOF-MS | [84] |
| | flavonoids, lipids, and alkaloids | OTEC-QTOT-MS | [04] |
| | Amino acids, peptides, and analogs; carbohydrates; | | |
| Dry-fermented | organic acids and derivatives; nucleosides, | II IID MACNIMD | F0.51 |
| sausages | nucleotides, and analogues; fatty acids and | H-HR-MAS NMR | [85] |
| | miscellaneous | | |
| G 1: | Amino acids, organic acids, aldoses, alditol, and | 10010 | 50.63 |
| Sunki | alcohol | NMR- and GC-MS | [86] |
| | Glycerophospholipids, fatty acyls, sphingolipids, 1- | | |
| Koumiss | glycerolipids, prenol lipids, organic acids, derivatives, | | |
| | organic oxygen, organoheterocyclic, benzenoids, | | |
| | organic nitrogen compounds, phenylpropanoids, | | |
| | polyketides, alkaloids, glycerophospholipids, fatty | UPLC-QTOF-MS | [87] |
| | acyls, included amino acids, carboxylic acids and | | |
| | derivatives, benzenoids, glycosides, glycerolipids, | | |
| | | | |
| | alcohols, lactones, carbonyl compounds. | | |

3. Probiotic Fermentation of Food Increases Bioactive Compounds

The bioactive compounds rich in plants, dairy products, and microorganisms have a wide range of biological potential. For example, antidiabetic, immunity regulation, anticancer agents [2,88,89], and antiviral [3,13,23]. The functionality of the bioactive compounds is based on their structure. The triple helical β -glucans having a low molecular weight and higher m stiffness showed strong anti-tumor ability as compared to the lower and higher molecular weight and stiffness β -glucans [90]. Similarly, owing to the higher molecular weight (~10 MDa) of the β -(1-4)-D-mannans, they have stronger immune stimulation potential than lower molecular weight (~1.3 MDa) [91]. Fermentation of food with probiotic bacteria enhances its nutritional values, modifies chemical structure, and increases the availability of bioactive compounds compared to the unfermented and traditionally fermented products (Table 3). In plant cell walls, components such as cellulose, hemicellulose, and lignin form a compact structure that is difficult to hydrolyze. However, during fermentation, probiotic bacteria help break down these complex substances into simpler subunits, thereby increasing the levels of bioactive compounds [65]. The rhamnogalacturonan-I-type polysaccharides were degraded in the process of probiotic fermentation in the carrot pulp, which increased their biological function than unfermented carrot [92]. *L. casei, Enterococcus faecalis*, and *Candida utilis* were used in the fermentation of the *Semen vaccariae* and *Leonurus artemisia* Chinese herbal medicine, which increased 55.14%, 127.28%, 55.42%, and 49.21%, respectively, of the total flavonoids, alkaloids,

polysaccharides, and saponins, compared with the natural herbs [93]. During the fermentation process of *Lespedeza cuneata* with *Lactobacillus pentosus*, the levels of quercetin and kaempferol were significantly increased by 242.9% and 266.7%, respectively. This enhancement augments the potential anti-aging properties of the fermented product [94].

The active regulation and metabolism of glucose and lipids in the probiotic-fermented carrot (Daucus carota L.) pulp enhance its functionality against diabetic effects compared to the unfermented pulp. It was also stated that polysaccharides of the probiotic-fermented pulp are more effective for type II diabetic rats than unfermented carrot pulp. This indicates that probiotic fermentation alters both the structure and concentration of bioactive compounds. When longan pulp is fermented by L. fermentum, the resulting polysaccharides exhibit characteristics such as lower molecular weight, viscosity, and particle size, yet higher solubility compared to those found in unfermented longan pulp. Therefore, Leuconostoc mesenteroides and L. casei were restored in the fermented longan pulp, which stimulates macrophage secretion of NO and IL-6 [95]. This indicates that the probiotic fermented food can lead to modification and enhancement in the physicochemical structure of the bioactivity of bioactive compounds. Additionally, probiotic fermentation extends the shelf life of foods and enhances their nutritional value. For example, Bacteroides spp could alter the structures of β-glucans, and glycans degraded into tri- and tetrasaccharides by L. plantarum [96]. The conversion of the baicalin to baicalein via its β -glucuronidase by L. brevis. Saccharomyces boulardii and L. plantarum bacteria tea fermentation were observed to improve the bioactive constituents, including methyl salicylate, geraniol, and 2-phenylethyl alcohol [97]. In the barley beverages, the contents of the total polyphenols and flavonoids were improved through fermention with strain L. casei and increased the antioxidant inhibitory function compared to the unfermented barley beverage [98]. Similarly, the lily bulb extract of L. lancifolium was cultivated with S. cerevisae, which increased polysaccharide production and the protein removal ratio compared to the unfermented. The carrot water-soluble and soybean-soluble polysaccharides, after the probiotic fermentation, enhanced immune regulation function [12]. Lily bulb fermented with S. cerevisiae showed a 91.46% protein removal ratio, while L. plantarum decreased the water-soluble polysaccharides by degrading them into monosaccharides. Puerariae radix was fermented with the B. breve increased 785% and 1010% of the daidzein and genistein contents which promoted the production of hyaluronic acid in the NHEK cells [99]. The incorporation of probiotic bacteria in food fermentation can reduce or change the toxic effects of the food.

Table 3. Probiotic fermentation of plant-based food increases bioactive compounds.

| Fermented Agent | Probiotic Strains | Increase Compounds | References |
|---|---|--|------------|
| Carrot pulp | L. plantarum NCU 116 | Rhamnogalacturonan-I-type polysaccharides break down | [92] |
| Semen vaccariae and Leonurus artemisia | L. casei, Enterococcus faecalis, and Candida utilis | Total flavonoids, alkaloids, crude polysaccharides, and saponins contents | [100] |
| Lespedeza cuneata | L. pentosus | Quercetin and kaempferol | [101] |
| Daucus carota L. | L. plantarum NCU116 | Effective regulation of glucose and lipid metabolism | [102] |
| Longan pulp | L. fermentum | Lower polysaccharide molecular weight, viscosity, and particle size, but higher solubility | [95] |
| Lily bulbs | L. plantarum | β-glucans and glycans degraded into tri- and tetra-saccharides | [103] |
| Lily bulbs | Limosilactobacillus fermentum GR-3 | hexadecanoic acid methyl ester, 22- tetrahydroxy-5alpha-cholestan-6-one-3-O- beta-D-allopyranoside, 22-O-(6-deoxy- Alpha-L-mannopyranosyl)-3-O-beta-D- glucopyranosylpregn-5-en-20-one, 1-O- trans-feruloylglycerol, and 3,4 dihydroxybenzoic acid | [4] |
| Tea Plant | Saccharomyces boulardii and L. plantarum | methyl salicylate, geraniol, and 2-phenyl ethyl alcohol | [104] |
| Barley beverage | L. casei | total polyphenols and flavonoid contents | [98] |
| Lily bulbs | L. lancifolium and S. cerevisae | protein contents | [105] |
| Puerariae radix | Bifidobactericum breve | daidzein and genistein | [106] |
| Soybean | Bacillus licheniformis | cheniformis Insulin-sensitizing action | |
| Artemisia princeps | L. plantarum SN13T | catechol and seco-tanapartholide C | [108] |

| Panax notoginseng | Streptococcus salivarius, L. helveticus, L.rhamnosus L.acidophilus, B. longum, B. catenulatum, B. breve and B. bifidum | ginsenosides Rh (1) and Rg (3) | [109] |
|-------------------------|--|--|-------|
| Polygonum cuspidatum | Aspergillus niger and Yeast | Production of resveratrol | [110] |
| Radix astragalus | Aspergillus spp. | 3,4-di(4'-hydroxyphenyl) isobutyric acid | [111] |
| Cordyceps militaris | Pediococcus pentosaceus | β-glucan and cordycepin | [112] |

4. Antiviral Properties of Fermented Food

Fermented foods and probiotic bacterial strains have received great attention for having the ability of high antiviral potential. The consumption of fermented food promotes the T-lymphocytes (CD³+, CD¹6+, and CD⁵6+) function and enhances pro-inflammatory cytokines to reduce the toxicity of the natural killer cells [113]. These functions of fermented food play a crucial role in controlling viral infections in both the alimentary and respiratory tracts. Furthermore, probiotic fermented foods support the digestive system and offer protection against influenza A viral infection [2]. Additionally, gastrointestinal viruses like rotaviruses, noroviruses, and enteroviruses can be controlled or minimized through the consumption of functional fermented foods [13]. Although the antiviral action of fermented products is not fully understood, studies have postulated that they may enhance the host's immunity through both direct and indirect viral contact (Figure 1). This mechanism showed that stimulation of the dendritic cells was increased because of the IgA secretion and IL-6 and IL-10 cells in the clinical trials by some probiotic strains from fermented food. IgA inhibition was caused to control the mucosal membrane attachment and attack of viruses [3]. Similarly, the lipopolysaccharides (LPS) prevent the viral ambition to the host cells. The detailed mechanism is presented in Table 4. It was found to inhibit influenza replication. The investigators stated that fermented foods comprising probiotics and bioactive compounds boost immune function and decrease casualty and viral infections.

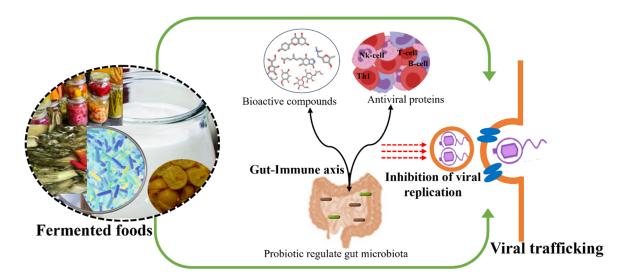


Figure 1. Fermented food promotes the immune system against viral infection via direct pathways (bioactive compounds such as organic acid, fatty acid, phenolic, and flavonoids) and indirectly by probiotics and gut microbiota.

4.1. Inhibition of Respiratory Virus

The influenza virus is a major cause of morbidity and mortality worldwide, primarily through respiratory tract infections. Emerging evidence suggests that the consumption of fermented vegetables, dairy products, and probiotic strains derived from fermented foods may help mitigate the effects of influenza. These dietary components have demonstrated the ability to enhance immune responses and may contribute to reducing the severity and duration of influenza infections. Therefore, incorporating fermented foods and probiotics into the diet represents a promising adjunct strategy for supporting immune defense against influenza. Thus, integrating fermented foods and probiotics into one's diet may offer a proactive approach to combating influenza-related health risks. In mice experiments, it was found that the influenza virus infections were controlled following

administration of *Lactobacillus*, thereby increasing survival rates of the mice[22]. Similarly, Kimchi has a rich diversity of *L. caseai* Dk128 was employed in the mice experiment to control or protect against the influenza virus infection by quickly starting antibodies against IgG 1 and IgG2. As well as inducing the immune function of innate immune cells and cytokines [38]. This data indicates that the dose-dependent probiotic bacterial administration in fermented foods could also affect viral inhibition. Additionally, Korean fermented cabbage was screened with the *L. plantarum* DK119 to increase the cytokines IL-12 and IFN-c levels while decreasing the inflammation in the bronchoalveolar lavage fluids infected by the influenza virus [114]. *L. bulgaricus* OLL1073R-1 fermented yogurt was used by the elderly (67-74 years) of people, which can reduce the common cold by controlling the natural cell killers and protecting the immune system [115]. These findings indicate that incorporating probiotic-rich fermented foods into the diet could serve as a valuable complementary strategy to enhance antiviral defense, particularly against influenza. However, further clinical studies are warranted to confirm these effects in human populations and to determine optimal strains and dosages for therapeutic or preventive use.

4.2 Gastrointestinal Virus Suppression

Fermented foods have shown promising potential in suppressing gastrointestinal viral infections through multiple mechanisms. Probiotic strains commonly found in fermented products, such as *Lactobacillus*, *Bifidobacterium*, and *Saccharomyces* species, contribute to strengthening the intestinal barrier, modulating the gut microbiota, and enhancing mucosal immunity. These beneficial microbes can inhibit viral adhesion to intestinal epithelial cells, compete for nutrients and receptor sites, and produce antiviral metabolites such as lactic acid, short-chain fatty acids (SCFAs), and bacteriocins. Additionally, it was stated that the use of fermented food inhibited the rotaviruses, noroviruses, and enteroviruses [77]. However, before attachment and fusion with host cells, viruses can be affected by various physiological barriers, such as proteolytic enzymes, bile salts, and low pH conditions in the gastrointestinal tract [116]. Nonetheless, some enteric viruses exhibit resistance to these harsh conditions. Interestingly, in the early stages of infection, these viruses may be disrupted by fermented foods enriched with probiotic-derived bioactive compounds [113]. Strains *L. ruminis* and *B. longum* SPM1205, and SPM1206 were observed in the fermented food to control the rotavirus infection by inhibiting the Caco-2 cells. The replication of the murine norovirus-1 (MNV-1) was inhibited by the co-culture of *Bifidobacterium adolescentis* and the RAW 264.7 cells [117].

4.2. Herpes Simplex Virus

HSV-2 and HSV-1 were most frequently developed in the body, which causes herpes infection [118]. A group of probiotic strains, including *L. lactis* subspecies *lactis*, *L. rhamnosus*, *L. brevis*, and *L. crispatus*, respectively, can inhibit HSV-1 and HSV-2 replication. Similarly, bacteriocins purified from the *L. lactis* association with the cell-surface bacterial component and heat resistance repressed HSV-1 and HSV-2 activities [119]. In mammalian Vero and HeLa cell lines, the inhibitory activities of strain *L. crispatus* were investigated against HSV-2. The results showed that the viral entry was evaded due to the blockage of the HSv-2 in the cell surface during the colonization of *L. crispatus* [120] (Table 3)

4.3. COVID-19

The COVID-19 outbreak has gained serious critical significance, not only in the form of casualties but also caused financial crises worldwide back 2020-2021 and still [121]. It presents seasonal epidemics ranging from mild illness to fatal pulmonary diseases, which have historically been combated through vaccination to some extent and antiviral treatments. Nonetheless, these treatments commonly have limited effectiveness in persons with immunity suppression. Therefore, an appropriate strategy is needed to investigate the natural compounds or agents that suppress SARS-CoV-2 activity and promote the immune system because presently available antiviral agents are not efficacious and might have health side effects. Fermented food and food additives rich in natural compounds may boost the immune function and control the COVID-19 infection. In such cases, for example, it was reported that fermented milk product used on a daily basis is a capable choice to treat various viral infections [121]. The fermented foods are rich in the probiotic community-related Lactobacilli genus, which was found to be highly effective against viral infection (Table 3). The modification of the gut microbiota promotes the immune function in COVID-19-infected patients. The probiotics control the pathogenic communities over the friendly bacteria despite their age [21], because, in the COVID-19-infected individual, the pathogenic microbes stress the immune function. Therefore, the utilization of fermented foods and probiotics can help to minimize stress on the immune system, thereby enhancing its anti-viral capabilities. Kefir has gained considerable commercial interest due to its content of health-promoting probiotics and bioactive compounds that support gut microbiota balance

and enhance immune function. For example, kefir consumption has been shown to stimulate the production of immune cells such as CD⁴⁺, CD⁸⁺, IgG⁺, IgA⁺, B, and T-cells, along with cytokines IL-2, IL-12, -γ. These immune components play critical roles in modulating immune responses and have been implicated in controlling cytokine storms during COVID-19 and other viral infections [22,52]. It has been proposed that incorporating probiotic bacterial strains into fermented foods to improve the bioavailability of beneficial compounds may offer a promising and supportive approach to the clinical management of COVID-19.

Table 4. Probiotic strains and bioactive compounds isolated from fermented food are effective against various kinds of viral infections.

| Antiviral Agent | Target Viruses | References |
|---|--|--------------|
| P. pentosaceus, W. cibaria, B. adolescentis | Noroviruses and, murine-1virus | [18,122] |
| L. brevis and Secoiridoid glucosides | Herpes simplex virus type 2 | [120] |
| L. acidophilus, L. acidophilus, L. rhamnosus, L. plantarum, S. thermophilus, and B. bifidu | Hepatitis C, Influenza virus, | [123,124] |
| L. acidophilus, L. reuteri, and L. salivarius | avian influenza virus | [125] |
| L. plantarum, Enterococcus faecium L3 | Influenza virus, Coxsackie virus, Echovirus E7, and E19 | [126] |
| Lactobacillus gasseri | Influenza A virus, Respiratory syncytial virus | [127] |
| L. reuteri ATCC 55730 | Coxsackieviruses CA6 and Enterovirus 71 | [128] |
| L. plantarum YU | Influenza A virus | [129] |
| L. plantarum DK119, L. gasseri SBT2055, L. casei DK128, Caffeic acid and glycyrrhizin Influenza virus | | [38,130,131] |
| Lactobacillus spp, Bifidobacteria | Vesicular stomatitis virus | [132,133] |
| Vitamin A, C, D, omega-3, fatty acids, and docosahexaenoic acid | Influenza and COVID-19 | [134,135] |
| Taurine, creatine, carnosine, anserine, and 4-hydroxyproline | COVID-19 | [136] |
| hexadecanoic acid methyl ester, 22-tetrahydroxy-5alpha-cholestan-6-one-3-O-beta-D-allopyranoside, | | |
| 22-O-(6-deoxy-Alpha-L-mannopyranosyl)-3-O-beta- D-glucopyranosylpregn-5-en-20-one, 1-O-trans- feruloylglycerol, and 3,4 dihydroxybenzoic acid | Lung infection healing | [4] |

5. Fermented Food Safety, Conclusion, and Future Prospects

The migration of people to urban areas has introduced several changes, including the need for food organization. Therefore, it is crucial to explore strategies for ensuring food security, in light of some outbreaks in recent years [31]. Several factors affect food security, including optimization parameters, economic pressure, and specific training for fermentation practices [137]. Among many food products, fermented food is highly produced at a small scale, and is considered tasty worldwide, although it may carry some risk. There are 3500 various types of fermented foods worldwide, accounting for up to 7.2% of the beverages in the period of 2018–2020 [1,138]. To boost consumer interest, attract entrepreneurs, and meet growing market demand, it's imperative to develop innovative approaches and product formulations in the realm of fermented foods. This might increase public demand and willingness to pay more attention to the use of fermented food and beverages such as grains, legumes, fruits, and vegetables. Fermented food product integration is important in national and global markets to sustain food care and production. Implementation of protective techniques can help reduce food-associated risks to public health, support free trade, improve food safety, and quality. Several probiotic strains can improve bioactive compounds and restore the abundance of beneficial microbiota in fermented food.

Furthermore, it is crucial to understand the dynamic interactions among metabolites and microbial diversity involved in the fermentation process to formulate microbiome models for assessing food quality and safety. Future research endeavors should explore and screen fermented products from diverse regions worldwide to identify probiotics and bioactive compounds that enhance immune system function. Additionally, fermented foods containing probiotics and bioactive compounds should be incorporated into clinical trials for viral infection management. Determining the optimal dose and duration of fermented food consumption is imperative. Nevertheless, the effects of bioactive compounds and probiotics present in fermented foods can enhance antiviral activities on host immune cells, both directly and indirectly. A significant challenge lies in the absence of an adequate database to profile microbiota and bioactive compounds in various fermented foods.

Author Contributions: X.Z.: Conceptualization, writing—original draft; B.C.: Formal analysis, writing—reviewing and editing; X.W.: Writing—reviewing and editing; A.K.: Conceptualization, writing—original draft; W.W.: Conceptualization, writing-reviewing and editing, Validation Resources, Project administration, Supervision. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable. **Data Availability Statement:** Not applicable.

Conflicts of Interest: The authors declare that there is no conflict of interest.

References

- 1. Lei, G.; Khan, A.; Budryn, G.; Grzelczyk, J. Probiotic products from laboratory to commercialization. *Trends Food Sci. Technol.* **2025**, *155*, 104807 https://doi.org/10.1016/j.tifs.2024.104807.
- Hoxha, R.; Todorov, D.; Hinkov, A.; Shishkova, K.; Evstatieva, Y.; Nikolova, D. In Vitro Screening of Antiviral Activity
 of Lactic Acid Bacteria Isolated from Traditional Fermented Foods. *Microbiol. Res.* 2023, 14, 333–342.
 https://doi.org/10.3390/microbiolres14010026.
- 3. Varsha, K.K.; Narisetty, V.; Brar, K.K.; Madhavan, A.; Alphy, M.P.; Sindhu, R.; Awasthi, M.K.; Varjani, S.; Binod, P. Bioactive metabolites in functional and fermented foods and their role as immunity booster and anti-viral innate mechanisms. *J. Food Sci. Technol.* **2022**, *60*, 2309–2318. https://doi.org/10.1007/s13197-022-05528-8.
- Khan, A.; Wang, W.; Ji, J.; Ling, Z.; Liu, P.; Xiao, S.; Han, H.; Salama, E.S.; Kumar Khanal, S.; Li, X. Fermented lily bulbs by Jiangshui probiotics improves lung health in mice. *Food Chem.* 2023, 440, 138270. https://doi.org/10.1016/j.foodchem.2023.13 8270.
- Intharuksa, A.; Kuljarusnont, S.; Sasaki, Y.; Tungmunnithum, D. Flavonoids and Other Polyphenols: Bioactive Molecules from Traditional Medicine Recipes/Medicinal Plants and Their Potential for Phytopharmaceutical and Medical Application. *Molecules* 2024, 29, 5760 https://doi.org/10.3390/molecules29235760.
- Özpınar, F.B.; İspirli, H.; Kayacan, S.; Korkmaz, K.; Dere, S.; Sagdic, O.; Alkay, Z.; Tunçil, Y.E.; Ayyash, M.; Dertli, E. Physicochemical and structural characterisation of a branched dextran type exopolysaccharide (EPS) from Weissella confusa S6 isolated from fermented sausage (Sucuk). *Int. J. Biol. Macromol.* 2024, 264, 130507. https://doi.org/10.1016/j.ijbiomac.2024.130507.
- 7. Zheng, Z.; Xie, G.; Liu, H.; Tan, G.; Li, L.; Liu, W.; Li, M. Fermented ginseng leaf enriched with rare ginsenosides relieves exercise-induced fatigue via regulating metabolites of muscular interstitial fluid, satellite cells-mediated muscle repair and gut microbiota. *J. Funct. Foods* **2021**, *83*, 104509. https://doi.org/10.1016/j.jff.2021.104509.
- 8. Zhu, C.; Guan, Q.; Song, C.; Zhong, L.; Ding, X.; Zeng, H.; Nie, P.; Song, L. Regulatory effects of Lactobacillus fermented black barley on intestinal microbiota of NAFLD rats. *Food Res. Int.* **2021**, *147*, 110467. https://doi.org/10.10 16/j.foodres.2021.110467.
- Zhao, Y.; Wu, C.; Zhu, Y.; Zhou, C.; Xiong, Z.; Samy Eweys, A.; Zhou, H.; Dong, Y.; Xiao, X. Metabolomics strategy for revealing the components in fermented barley extracts with Lactobacillus plantarum dy-1. *Food Res. Int.* 2021, 139, 109808. https://doi.org/10.1016/j.foodres.2020.109808.
- 10. Peruzzolo, M.; Ceni, G.C.; Junges, A.; Zeni, J.; Cansian, R.L.; Backes, G.T. Probiotics: Health benefits, microencapsulation, and viability, combination with natural compounds, and applications in foods. *Food Biosci.* **2025**, *66*, 106253. https://doi.org/10.1016/j.fbio.2025.106253.
- 11. Sakandar, H.A.; Zhang, H. Trends in Probiotic(s)-Fermented milks and their in vivo functionality: A review. *Trends Food Sci. Technol.* **2021**, *110*, 55–65. https://doi.org/10.1016/j.tifs.2021.01.054.
- 12. Muhialdin, B.J.; Kadum, H.; Meor Hussin, A.S. Metabolomics profiling of fermented cantaloupe juice and the potential application to extend the shelf life of fresh cantaloupe juice for six months at 8 °C. *Food Control* **2021**, *120*, 107555. https://doi.org/10.1016/j.foodcont.2020.107555.
- 13. Wan-Mohtar, W.A.A.Q.I.; Ilham, Z.; Jamaludin, A.A.; David, W.; Mohd Zaini, N.A. Fermented foods as alternative functional foods during post-pandemic in Asia. *Front. Food Sci. Technol.* **2022**, *2*, 1047970. https://doi.org/10.3389/frfst. 2022.1047970.
- 14. Wen-qiong, W.; Jie-long, Z.; Qian, Y.; Ji-yang, Z.; Mao-lin, L.; Rui-xia, G.; Yujun, H. Structural and compositional changes of whey protein and blueberry juice fermented using Lactobacillus plantarum or Lactobacillus casei during fermentation. *RSC Adv.* **2021**, *11*, 26291–26302. https://doi.org/10.1039/d1ra04140a.
- 15. Kayacan Çakmakoğlu, S.; Dere, S.; Bekiroğlu, H.; Bozkurt, F.; Karasu, S.; Dertli, E.; Türker, M.; Sagdic, O. Production of bioactive peptides during yogurt fermentation, their extraction and functional characterization. *Food Biosci.* **2024**, *61*, 104805. https://doi.org/10.1016/j.fbio.2024.104805.

- 16. Wei, J.; Zhang, Y.; Li, H.; Wang, F.; Yao, S. Toll-like receptor 4: A potential therapeutic target for multiple human diseases. *Biomed. Pharmacother.* **2023**, *166*, 115338. https://doi.org/10.1016/j.biopha.2023.115338.
- 17. de Araújo, F.F.; Farias, D.P. Psychobiotics: An emerging alternative to ensure mental health amid the COVID-19 outbreak? *Trends Food Sci. Technol.* **2020**, *103*, 386–387. https://doi.org/10.1016/j.tifs.2020.07.006.
- 18. Seo, D.J.; Jung, D.; Jung, S.; Yeo, D.; Choi, C. Inhibitory effect of lactic acid bacteria isolated from kimchi against murine norovirus. *Food Control* **2020**, *109*, 106881. https://doi.org/10.1016/j.foodcont.2019.106881.
- 19. Starosila, D.; Rybalko, S.; Varbanetz, L.; Ivanskaya, N.; Sorokulova, I. Anti-influenza Activity of a Bacillus subtilis Probiotic Strain. *Antimicrob. Agents Chemother.* **2017**, *61*, 00539–17. https://doi.org/10.1128/aac.00539-17.
- 20. Tonucci, L.B.; dos Santos Olbrich, K.M.; de Oliveira Licursi, L.; Rocha Ribeiro, S.M.; Duarte Martino, H.S. Clinical application of probiotics in type 2 diabetes mellitus: A randomized, double-blind, placebo-controlled study. *Clin. Nutr.* **2017**, *36*, 85–92. https://doi.org/10.1016/j.clnu.2015.11.011.
- 21. Mirashrafi, S.; Moravejolahkami, A.R.; Balouch Zehi, Z.; Hojjati Kermani, M.A.; Bahreini-Esfahani, N.; Haratian, M.; Ganjali Dashti, M.; Pourhossein, M. The efficacy of probiotics on virus titres and antibody production in virus diseases: A systematic review on recent evidence for COVID-19 treatment. *Clin. Nutr. ESPEN* **2021**, *46*, 1–8. https://doi.org/10.10 16/j.clnesp.2021.10.016.
- 22. Hamida, R.S.; Shami, A.; Ali, M.A.; Almohawes, Z.N.; Mohammed, A.E.; Bin-Meferij, M.M. Kefir: A protective dietary supplementation against viral infection. *Biomed. Pharmacother.* **2021**, *133*, 110974. https://doi.org/10.1016/j.biopha.20 20.110974.
- 23. Tomas, M.; Capanoglu, E.; Bahrami, A.; Hosseini, H.; Akbari-Alavijeh, S.; Shaddel, R.; Rehman, A.; Rezaei, A.; Rashidinejad, A.; Garavand, F.; et al. The direct and indirect effects of bioactive compounds against coronavirus. *Food Front.* **2021**, *3*, 96–123. https://doi.org/10.1002/fft2.119.
- 24. Wang, S.; Liu, X.; Tamura, T.; Kyouno, N.; Zhang, H.; Chen, J.Y. Effect of volatile compounds on the quality of miso (traditional Japanese fermented soybean paste). *Lwt* **2021**, *139*, 110573. https://doi.org/10.1016/j.lwt.2020.110573.
- 25. Wu, Y.; Ye, Z.; Feng, P.; Li, R.; Chen, X.; Tian, X.; Han, R.; Kakade, A.; Liu, P.; Li, X. Limosilactobacillus fermentum JL-3 isolated from "Jiangshui" ameliorates hyperuricemia by degrading uric acid. *Gut Microbes* **2021**, *13*, 1897211. https://doi.org/10.1080/19490976.2021.1897211.
- 26. Kim, N.; Lee, J.; Song, H.S.; Oh, Y.J.; Kwon, M.-S.; Yun, M.; Lim, S.K.; Park, H.K.; Jang, Y.S.; Lee, S.; et al. Kimchi intake alleviates obesity-induced neuroinflammation by modulating the gut-brain axis. *Food Res. Int.* **2022**, *158*, 111533. https://doi.org/10.1016/j.foodres.2022.111533.
- 27. Tamang, J.P.; Anupma, A.; Nakibapher Jones Shangpliang, H. Ethno-microbiology of Tempe, an Indonesian fungal-fermented soybean food and Koji, a Japanese fungal starter culture. *Curr. Opin. Food Sci.* **2022**, *48*, 100912. https://doi.org/10.1016/j.cofs.2022.100912.
- 28. Hinojosa-Avila, C.R.; García-Gamboa, R.; Chedraui-Urrea, J.J.T.; García-Cayuela, T. Exploring the potential of probiotic-enriched beer: Microorganisms, fermentation strategies, sensory attributes, and health implications. *Food Res. Int.* **2024**, *175*, 113717. https://doi.org/10.1016/j.foodres.2023.113717.
- 29. Grom, L.C.; Coutinho, N.M.; Guimarães, J.T.; Balthazar, C.F.; Silva, R.; Rocha, R.S.; Freitas, M.Q.; Duarte, M.C.K.H.; Pimentel, T.C.; Esmerino, E.A.; et al. Probiotic dairy foods and postprandial glycemia: A mini-review. *Trends Food Sci. Technol.* **2020**, *101*, 165–171. https://doi.org/10.1016/j.tifs.2020.05.012.
- 30. Rezac, S.; Kok, C.R.; Heermann, M.; Hutkins, R. Fermented Foods as a Dietary Source of Live Organisms. *Front. Microbiol.* **2018**, *9*, 1785. https://doi.org/10.3389/fmicb.2018.01785.
- 31. Zhang, T.; Geng, S.; Cheng, T.; Mao, K.; Chitrakar, B.; Gao, J.; Sang, Y. From the past to the future: Fermented milks and their health effects against human diseases. *Food Front.* **2023**, *4*, 1747–1777. https://doi.org/10.1002/fft2.304.
- 32. Miyamoto, M.; Ueno, H.M.; Watanabe, M.; Tatsuma, Y.; Seto, Y.; Miyamoto, T.; Nakajima, H. Distinctive proteolytic activity of cell envelope proteinase of Lactobacillus helveticus isolated from airag, a traditional Mongolian fermented mare's milk. *Int. J. Food Microbiol.* **2015**, *197*, 65–71. https://doi.org/10.1016/j.ijfoodmicro.2014.12.012.
- 33. Endo, A.; Irisawa, T.; Dicks, L.; Tanasupawat, S. Fermented Foods | Fermentations of East and Southeast Asia; Elsevier: Amsterdam, The Netherlands, 2014; pp. 846–851. https://doi.org/10.1016/b978-0-12-384730-0.00119-1.
- 34. Anandharaj, M.; Sivasankari, B.; Santhanakaruppu, R.; Manimaran, M.; Rani, R.P.; Sivakumar, S. Determining the probiotic potential of cholesterol-reducing Lactobacillus and Weissella strains isolated from gherkins (fermented cucumber) and south Indian fermented koozh. *Res. Microbiol.* **2015**, *166*, 428–439. https://doi.org/10.1016/j.resmic.2015.
- 35. Nielsen, B.; Gürakan, G.C.; Ünlü, G. Kefir: A Multifaceted Fermented Dairy Product. *Probiotics Antimicrob. Proteins* **2014**, *6*, 123–135. https://doi.org/10.1007/s12602-014-9168-0.
- 36. Wu, R.; Wang, L.; Wang, J.; Li, H.; Menghe, B.; Wu, J.; Guo, M.; Zhang, H. Isolation and preliminary probiotic selection of lactobacilli from koumiss in Inner Mongolia. *J. Basic Microbiol.* **2009**, *49*, 318–326. https://doi.org/10.1002/jobm.200 800047.

- 37. Xiong, T.; Guan, Q.; Song, S.; Hao, M.; Xie, M. Dynamic changes of lactic acid bacteria flora during Chinese sauerkraut fermentation. *Food Control* **2012**, *26*, 178–181. https://doi.org/10.1016/j.foodcont.2012.01.027.
- 38. Jung, Y.-J.; Lee, Y.-T.; Ngo, V.L.; Cho, Y.-H.; Ko, E.-J.; Hong, S.-M.; Kim, K.-H.; Jang, J.-H.; Oh, J.-S.; Park, M.-K.; et al. Heat-killed Lactobacillus casei confers broad protection against influenza A virus primary infection and develops heterosubtypic immunity against future secondary infection. *Sci. Rep.* **2017**, *7*, 17360 https://doi.org/10.1038/s41598-017-17487-8.
- 39. Xu, H.; Wang, J.; Liu, H.; Shen, G.; Zhang, Z.; Mei, X.; Sun, Y.; Dong, Y.; Wei, D.; Wang, W. Screening of uric acid-degrading lactic acid bacteria consortium and strain combination for enhancing degradation. *Microbiol. China* **2023**, *50*, 2545–2555. https://doi.org/10.13344/j.microbiol.china.220830.
- Liu, C.-J.; Tang, X.-D.; Yu, J.; Zhang, H.-Y.; Li, X.-R. Gut microbiota alterations from different Lactobacillus probioticfermented yoghurt treatments in slow-transit constipation. *J. Funct. Foods* 2017, 38, 110–118. https://doi.org/10.1016/j. iff.2017.08.037.
- 41. Anzawa, D.; Mawatari, T.; Tanaka, Y.; Yamamoto, M.; Genda, T.; Takahashi, S.; Nishijima, T.; Kamasaka, H.; Suzuki, S.; Kuriki, T. Effects of synbiotics containing *Bifidobacterium animalis* subsp. lactis GCL2505 and inulin on intestinal bifidobacteria: A randomized, placebo-controlled, crossover study. *Food Sci. Nutr.* **2019**, *7*, 1828–1837.
- 42. Akhtar, S.; Hussain, M.; Ismail, T.; Riaz, M. *Ethnic Fermented Foods of Pakistan*; Springer: New Delhi, India, 2016; pp. 119–137. https://doi.org/10.1007/978-81-322-2800-4 5.
- 43. Sakandar, H.A.; Usman, K.; Imran, M. Isolation and characterization of gluten-degrading Enterococcus mundtii and Wickerhamomyces anomalus, potential probiotic strains from indigenously fermented sourdough (Khamir). *Lwt* **2018**, 91, 271–277. https://doi.org/10.1016/j.lwt.2018.01.023.
- 44. Khan, A.; Li, S.; Han, H.; Jin, W.-L.; Ling, Z.; Ji, J.; Iram, S.; Liu, P.; Xiao, S.; Salama, E.-S.; et al. A gluten degrading probiotics relieves CeD symptom and normalize immune system in mice A gluten degrading probiotic Bacillus subtilis LZU-GM relieve adverse effect of gluten additive food and balances gut microbiota in mice. *Food Res. Int.* **2023**, *170*, 112960. https://doi.org/10.1016/j.foodres.2023.112960.
- 45. Nahidul-Islam, S.M.; Kuda, T.; Takahashi, H.; Kimura, B. Bacterial and fungal microbiota in traditional Bangladeshi fermented milk products analysed by culture-dependent and culture-independent methods. *Food Res. Int.* **2018**, *111*, 431–437. https://doi.org/10.1016/j.foodres.2018.05.048.
- 46. Zhadyra, S.; Han, X.; Anapiyayev, B.B.; Tao, F.; Xu, P. Bacterial diversity analysis in Kazakh fermented milks Shubat and Ayran by combining culture-dependent and culture-independent methods. *Lwt* **2021**, *141*, 110877. https://doi.org/10.1016/j.lwt.2021.110877.
- 47. Peres, C.M.; Alves, M.; Hernandez-Mendoza, A.; Moreira, L.; Silva, S.; Bronze, M.R.; Vilas-Boas, L.; Peres, C.; Malcata, F.X. Novel isolates of lactobacilli from fermented Portuguese olive as potential probiotics. *LWT Food Sci. Technol.* **2014**, 59, 234–246. https://doi.org/10.1016/j.lwt.2014.03.003.
- 48. Abriouel, H.; Omar, N.B.; López, R.L.; Martínez-Cañamero, M.; Keleke, S.; Gálvez, A. Culture-independent analysis of the microbial composition of the African traditional fermented foods poto poto and dégué by using three different DNA extraction methods. *Int. J. Food Microbiol.* **2006**, *111*, 228–233. https://doi.org/10.1016/j.ijfoodmicro.2006.06.006.
- Liang, T.; Xie, X.; Zhang, J.; Ding, Y.; Wu, Q. Bacterial community and composition of different traditional fermented dairy products in China, South Africa, and Sri Lanka by high-throughput sequencing of 16S rRNA genes. *Lwt* 2021, *144*, 111209. https://doi.org/10.1016/j.lwt.2021.111209.
- 50. Shori, A.B. Camel milk and its fermented products as a source of potential probiotic strains and novel food cultures: A mini review. *PharmaNutrition* **2017**, *5*, 84–88. https://doi.org/10.1016/j.phanu.2017.06.003.
- 51. Sengun, I.Y.; Nielsen, D.S.; Karapinar, M.; Jakobsen, M. Identification of lactic acid bacteria isolated from Tarhana, a traditional Turkish fermented food. *Int. J. Food Microbiol.* **2009**, *135*, 105–111. https://doi.org/10.1016/j.ijfoodmicro.20 09 07 033
- 52. Das, G.; Heredia, J.B.; de Lourdes Pereira, M.; Coy-Barrera, E.; Rodrigues Oliveira, S.M.; Gutiérrez-Grijalva, E.P.; Cabanillas-Bojórquez, L.A.; Shin, H.-S.; Patra, J.K. Korean traditional foods as antiviral and respiratory disease prevention and treatments: A detailed review. *Trends Food Sci. Technol.* **2021**, *116*, 415–433. https://doi.org/10.1016/j. tifs.2021.07.037.
- 53. Patel, A.; Prajapati, J.B.; Holst, O.; Ljungh, A. Determining probiotic potential of exopolysaccharide producing lactic acid bacteria isolated from vegetables and traditional Indian fermented food products. *Food Biosci.* **2014**, *5*, 27–33. https://doi.org/10.1016/j.fbio.2013.10.002.
- 54. Şanlier, N.; Gökcen, B.B.; Sezgin, A.C. Health benefits of fermented foods. *Crit. Rev. Food Sci. Nutr.* **2017**, *59*, 506–527. https://doi.org/10.1080/10408398.2017.1383355.
- 55. Santiago-López, L.; Aguilar-Toalá, J.E.; Hernández-Mendoza, A.; Vallejo-Cordoba, B.; Liceaga, A.M.; González-Córdova, A.F. Invited review: Bioactive compounds produced during cheese ripening and health effects associated with aged cheese consumption. *J. Dairy Sci.* **2018**, *101*, 3742–3757. https://doi.org/10.3168/jds.2017-13465.

- 56. Quigley, L.; O'Sullivan, O.; Beresford, T.P.; Ross, R.P.; Fitzgerald, G.F.; Cotter, P.D. Molecular approaches to analysing the microbial composition of raw milk and raw milk cheese. *Int. J. Food Microbiol.* **2011**, *150*, 81–94. https://doi.org/10.1016/j.ijfoodmicro.2011.08.001.
- 57. Saithong, P.; Panthavee, W.; Boonyaratanakornkit, M.; Sikkhamondhol, C. Use of a starter culture of lactic acid bacteria in plaa-som, a Thai fermented fish. *J. Biosci. Bioeng.* **2010**, *110*, 553–557. https://doi.org/10.1016/j.jbiosc.2010.06.004.
- 58. Barus, T.; Kristani, A.; Yulandi, A.D.I. Diversity of Amylase-Producing Bacillus spp. from "Tape" (Fermented Cassava). *HAYATI J. Biosci.* **2013**, *20*, 94–98. https://doi.org/10.4308/hjb.20.2.94.
- 59. Raji, M.N.A.; Ab Karim, S.; Ishak, F.A.C.; Arshad, M.M. Past and present practices of the Malay food heritage and culture in Malaysia. *J. Ethn. Foods* **2017**, *4*, 221–231. https://doi.org/10.1016/j.jef.2017.11.001.
- 60. Chettri, R.; Tamang, J.P. Bacillus species isolated from tungrymbai and bekang, naturally fermented soybean foods of India. *Int. J. Food Microbiol.* **2015**, *197*, 72–76. https://doi.org/10.1016/j.ijfoodmicro.2014.12.021.
- 61. Mishra, B.K.; Hati, S.; Das, S. Bio-nutritional aspects of Tungrymbai, an ethnic functional fermented soy food of Khasi Hills, Meghalaya, India. *Clin. Nutr. Exp.* **2019**, *26*, 8–22. https://doi.org/10.1016/j.yclnex.2019.05.004.
- 62. Dajanta, K.; Apichartsrangkoon, A.; Chukeatirote, E.; Frazier, R.A. Free-amino acid profiles of thua nao, a Thai fermented soybean. *Food Chem.* **2011**, *125*, 342–347. https://doi.org/10.1016/j.foodchem.2010.09.002.
- 63. Mahidsanan, T.; Gasaluck, P.; Eumkeb, G. A novel soybean flour as a cryoprotectant in freeze-dried Bacillus subtilis SB-MYP-1. *Lwt* **2017**, 77, 152–159. https://doi.org/10.1016/j.lwt.2016.11.015.
- 64. Cai, H.; Dumba, T.; Sheng, Y.; Li, J.; Lu, Q.; Liu, C.; Cai, C.; Feng, F.; Zhao, M. Microbial diversity and chemical property analyses of sufu products with different producing regions and dressing flavors. *Lwt* **2021**, *144*, 111245. https://doi.org/10.1016/j.lwt.2021.111245.
- 65. Yao, D.; Xu, L.; Wu, M.; Wang, X.; Zhu, L.; Wang, C. Effects of microbial community succession on flavor compounds and physicochemical properties during CS sufu fermentation. *Lwt* **2021**, *152*, 112313. https://doi.org/10.1016/j.lwt.2021. 112313.
- Dank, A.; van Mastrigt, O.; Yang, Z.; Dinesh, V.M.; Lillevang, S.K.; Weij, C.; Smid, E.J. The cross-over fermentation concept and its application in a novel food product: The dairy miso case study. *Lwt* 2021, *142*, 111041. https://doi.org/10.1016/j.lwt.2021. 111041.
- 67. Panda, A.; Ghosh, K.; Ray, M.; Nandi, S.K.; Parua, S.; Bera, D.; Singh, S.N.; Dwivedi, S.K.; Mondal, K.C. Ethnic preparation and quality assessment of Chhurpi, a home-made cheese of Ladakh, India. *J. Ethn. Foods* **2016**, *3*, 257–262. https://doi.org/10.1016/j.jef.2016.12.004.
- 68. Rai, A.K.; Kumari, R.; Sanjukta, S.; Sahoo, D. Production of bioactive protein hydrolysate using the yeasts isolated from soft chhurpi. *Bioresour. Technol.* **2016**, *219*, 239–245. https://doi.org/10.1016/j.biorech.2016.07.129.
- 69. Dewan, S.; Tamang, J.P. Dominant lactic acid bacteria and their technological properties isolated from the Himalayan ethnic fermented milk products. *Antonie Van Leeuwenhoek* **2007**, *92*, 343–352. https://doi.org/10.1007/s10482-007-9163-5.
- 70. Todorov, S.D. Diversity of bacteriocinogenic lactic acid bacteria isolated from boza, a cereal-based fermented beverage from Bulgaria. *Food Control* **2010**, *21*, 1011–1021. https://doi.org/10.1016/j.foodcont.2009.12.020.
- 71. Yeğin, S.; Üren, A. Biogenic amine content of boza: A traditional cereal-based, fermented Turkish beverage. *Food Chem.* **2008**, *111*, 983–987. https://doi.org/10.1016/j.foodchem.2008.05.020.
- 72. Chao, S.-H.; Wu, R.-J.; Watanabe, K.; Tsai, Y.-C. Diversity of lactic acid bacteria in suan-tsai and fu-tsai, traditional fermented mustard products of Taiwan. *Int. J. Food Microbiol.* **2009**, *135*, 203–210. https://doi.org/10.1016/j.ijfoodmicro. 2009.07.032.
- 73. Nguyen, D.T.L.; Van Hoorde, K.; Cnockaert, M.; De Brandt, E.; De Bruyne, K.; Le, B.T.; Vandamme, P. A culture-dependent and -independent approach for the identification of lactic acid bacteria associated with the production of nem chua, a Vietnamese fermented meat product. *Food Res. Int.* **2013**, *50*, 232–240. https://doi.org/10.1016/j.foodres.2012.09.029.
- 74. Tomita, S.; Watanabe, J.; Kuribayashi, T.; Tanaka, S.; Kawahara, T. Metabolomic evaluation of different starter culture effects on water-soluble and volatile compound profiles in nozawana pickle fermentation. *Food Chem. Mol. Sci.* **2021**, 2, 100019. https://doi.org/10.1016/j.fochms.2021.100019.
- 75. Daliri, E.B.-M.; Tyagi, A.; Ofosu, F.K.; Chelliah, R.; Kim, J.-H.; Kim, J.-R.; Yoo, D.; Oh, D.-H. A discovery-based metabolomic approach using UHPLC Q-TOF MS/MS unveils a plethora of prospective antihypertensive compounds in Korean fermented soybeans. *Lwt* **2021**, *137*, 110399. https://doi.org/10.1016/j.lwt.2020.110399.
- Zha, M.; Li, K.; Zhang, W.; Sun, Z.; Kwok, L.-Y.; Menghe, B.; Chen, Y. Untargeted mass spectrometry-based metabolomics approach unveils molecular changes in milk fermented by Lactobacillus plantarum P9. Lwt 2021, 140, 110759. https://doi.org/10.1016/j.lwt.2020.110759.
- 77. Muhialdin, B.J.; Zawawi, N.; Abdull Razis, A.F.; Bakar, J.; Zarei, M. Antiviral activity of fermented foods and their probiotics bacteria towards respiratory and alimentary tracts viruses. *Food Control* **2021**, *127*, 108140. https://doi.org/10.1016/j.foodcont.2021.108140.

- 78. Chan, M.Z.A.; Lau, H.; Lim, S.Y.; Li, S.F.Y.; Liu, S.-Q. Untargeted LC-QTOF-MS/MS based metabolomics approach for revealing bioactive components in probiotic fermented coffee brews. *Food Res. Int.* **2021**, *149*, 110656. https://doi.org/10.1016/j.foodres.2021.110656.
- Ayyash, M.; Abdalla, A.; Alhammadi, A.; Senaka Ranadheera, C.; Affan Baig, M.; Al-Ramadi, B.; Chen, G.; Kamal-Eldin, A.; Huppertz, T. Probiotic survival, biological functionality and untargeted metabolomics of the bioaccessible compounds in fermented camel and bovine milk after in vitro digestion. *Food Chem.* 2021, 363, 130243. https://doi.org/10.1016/j.foodchem.2021.130243.
- 80. Jia, W.; Liu, Y.; Shi, L. Integrated metabolomics and lipidomics profiling reveals beneficial changes in sensory quality of brown fermented goat milk. *Food Chem.* **2021**, *364*, 130378. https://doi.org/10.1016/j.foodchem.2021.130378.
- 81. Kang, H.J.; Yang, H.J.; Kim, M.J.; Han, E.-S.; Kim, H.-J.; Kwon, D.Y. Metabolomic analysis of meju during fermentation by ultra performance liquid chromatography-quadrupole-time of flight mass spectrometry (UPLC-Q-TOF MS). *Food Chem.* **2011**, *127*, 1056–1064. https://doi.org/10.1016/j.foodchem.2011.01.080.
- 82. Ferri, M.; Serrazanetti, D.I.; Tassoni, A.; Baldissarri, M.; Gianotti, A. Improving the functional and sensorial profile of cereal-based fermented foods by selecting Lactobacillus plantarum strains via a metabolomics approach. *Food Res. Int.* **2016**, *89*, 1095–1105. https://doi.org/10.1016/j.foodres.2016.08.044.
- 83. Gao, Y.X.; Xu, B.; Fan, H.R.; Zhang, M.R.; Zhang, L.J.; Lu, C.; Zhang, N.N.; Fan, B.; Wang, F.Z.; Li, S. 1H NMR-based chemometric metabolomics characterization of soymilk fermented by Bacillus subtilis BSNK-5. *Food Res. Int.* **2020**, *138*, 109686. https://doi.org/10.1016/j.foodres.2020.109686.
- 84. Lee, D.E.; Shin, G.R.; Lee, S.; Jang, E.S.; Shin, H.W.; Moon, B.S.; Lee, C.H. Metabolomics reveal that amino acids are the main contributors to antioxidant activity in wheat and rice gochujangs (Korean fermented red pepper paste). *Food Res. Int.* **2016**, *87*, 10–17. https://doi.org/10.1016/j.foodres.2016.06.015.
- 85. García-García, A.B.; Lamichhane, S.; Castejón, D.; Cambero, M.I.; Bertram, H.C. 1H HR-MAS NMR-based metabolomics analysis for dry-fermented sausage characterization. *Food Chem.* **2018**, *240*, 514–523. https://doi.org/10.1016/j.foodchem.2017. 07.150.
- 86. Tomita, S.; Nakamura, T.; Okada, S. NMR- and GC/MS-based metabolomic characterization of sunki, an unsalted fermented pickle of turnip leaves. *Food Chem.* **2018**, *258*, 25–34. https://doi.org/10.1016/j.foodchem.2018.03.038.
- 87. Xia, Y.; Yu, J.; Miao, W.; Shuang, Q. A UPLC-Q-TOF-MS-based metabolomics approach for the evaluation of fermented mare's milk to koumiss. *Food Chem.* **2020**, *320*, 126619. https://doi.org/10.1016/j.foodchem.2020.126619.
- 88. Qu, Q.; Yang, F.; Zhao, C.; Liu, X.; Yang, P.; Li, Z.; Han, L.; Shi, X. Effects of fermented ginseng on the gut microbiota and immunity of rats with antibiotic-associated diarrhea. *J. Ethnopharmacol.* **2021**, *267*, 113594. https://doi.org/10.1016/j.jep.2020.113594.
- 89. Ghyselinck, J.; Verstrepen, L.; Moens, F.; Van Den Abbeele, P.; Bruggeman, A.; Said, J.; Smith, B.; Barker, L.A.; Jordan, C.; Leta, V.; et al. Influence of probiotic bacteria on gut microbiota composition and gut wall function in an in-vitro model in patients with Parkinson's disease. *Int. J. Pharm. X* 2021, *3*, 100087. https://doi.org/10.1016/j.ijpx.2021.100087.
- 90. Banwo, K.; Olojede, A.O.; Adesulu-Dahunsi, A.T.; Verma, D.K.; Thakur, M.; Tripathy, S.; Singh, S.; Patel, A.R.; Gupta, A.K.; Aguilar, C.N.; et al. Functional importance of bioactive compounds of foods with Potential Health Benefits: A review on recent trends. *Food Biosci.* **2021**, *43*, 101320. https://doi.org/10.1016/j.fbio.2021.101320.
- 91. Jabbari, F.; Babaeipour, V.; Saharkhiz, S. Comprehensive review on biosynthesis of hyaluronic acid with different molecular weights and its biomedical applications. *Int. J. Biol. Macromol.* **2023**, *240*, 124484. https://doi.org/10.1016/j.ijbiomac.2023.124484.
- 92. Wan, Y.-J.; Hong, T.; Shi, H.-F.; Yin, J.-Y.; Koev, T.; Nie, S.-P.; Gilbert, R.G.; Xie, M.-Y. Probiotic fermentation modifies the structures of pectic polysaccharides from carrot pulp. *Carbohydr. Polym.* **2021**, *251*, 117116. https://doi.org/10.1016/j.carbpol.2020.117116.
- 93. Zhang, Y.; Zhang, J.; Yan, J.; Qi, X.; Wang, Y.; Zheng, Z.; Liang, J.; Ling, J.; Chen, Y.; Tang, X.; et al. Application of fermented Chinese herbal medicines in food and medicine field: From an antioxidant perspective. *Trends Food Sci. Technol.* **2024**, *148*, 104410. https://doi.org/10.1016/j.tifs.2024.104410.
- 94. Lee, K.S.; Park, S.N. Cytoprotective effects and mechanisms of quercetin, quercitrin and avicularin isolated from Lespedeza cuneata G. Don against ROS-induced cellular damage. *J. Ind. Eng. Chem.* **2019**, *71*, 160–166. https://doi.org/10.1016/j.jiec.2018.11.018.
- 95. Huang, F.; Hong, R.; Zhang, R.; Yi, Y.; Dong, L.; Liu, L.; Jia, X.; Ma, Y.; Zhang, M. Physicochemical and biological properties of longan pulp polysaccharides modified by Lactobacillus fermentum fermentation. *Int. J. Biol. Macromol.* **2019**, *125*, 232–237. https://doi.org/10.1016/j.ijbiomac.2018.12.061.
- 96. Fernandez-Julia, P.J.; Munoz-Munoz, J.; van Sinderen, D. A comprehensive review on the impact of β-glucan metabolism by Bacteroides and Bifidobacterium species as members of the gut microbiota. *Int. J. Biol. Macromol.* **2021**, *181*, 877–889. https://doi.org/10.1016/j.ijbiomac.2021.04.069.
- 97. Hu, T.; Shi, S.; Ma, Q. Modulation effects of microorganisms on tea in fermentation. *Front. Nutr.* **2022**, *9*, 931790. https://doi.org/10.3389/fnut.2022.931790.

- 98. Guo, W.; Chen, M.; Cui, S.; Tang, X.; Zhang, Q.; Zhao, J.; Mao, B.; Zhang, H. Effects of Lacticaseibacillus casei fermentation on the bioactive compounds, volatile and non-volatile compounds, and physiological properties of barley beverage. *Food Biosci.* **2023**, *53*, 102695. https://doi.org/10.1016/j.fbio.2023.102695.
- 99. Choi, Y.; Bose, S.; Shin, N.R.; Song, E.-J.; Nam, Y.-D.; Kim, H. Lactate-Fortified Puerariae Radix Fermented by Bifidobacterium breve Improved Diet-Induced Metabolic Dysregulation via Alteration of Gut Microbial Communities. *Nutrients* **2020**, *12*, 276. https://doi.org/10.3390/nu12020276.
- 100. Li, L.; Wang, L.; Fan, W.; Jiang, Y.; Zhang, C.; Li, J.; Peng, W.; Wu, C. The Application of Fermentation Technology in Traditional Chinese Medicine: A Review. *Am. J. Chin. Med.* 2020, 48, 899–921. https://doi.org/10.1142/s0192415x20500433.
- 101. Seong, J.S.; Xuan, S.H.; Park, S.H.; Lee, K.S.; Park, Y.M.; Park, S.N. Antioxidative and Antiaging Activities and Component Analysis of Lespedeza cuneata G. Don Extracts Fermented with Lactobacillus pentosus. *J. Microbiol. Biotechnol.* 2017, 27, 1961–1970. https://doi.org/10.4014/jmb.1706.06028.
- 102. Li, C.; Ding, Q.; Nie, S.-P.; Zhang, Y.-S.; Xiong, T.; Xie, M.-Y. Carrot Juice Fermented with Lactobacillus plantarum NCU116 Ameliorates Type 2 Diabetes in Rats. *J. Agric. Food Chem.* **2014**, *62*, 11884–11891. https://doi.org/10.1021/jf503681r.
- 103. Song, S.; Liu, X.; Zhao, B.; Abubaker, M.A.; Huang, Y.; Zhang, J. Effects of Lactobacillus plantarum Fermentation on the Chemical Structure and Antioxidant Activity of Polysaccharides from Bulbs of Lanzhou Lily. *ACS Omega* **2021**, *6*, 29839–29851. https://doi.org/10.1021/acsomega.1c04339.
- 104. Wang, R.; Sun, J.; Lassabliere, B.; Yu, B.; Liu, S.Q. Green tea fermentation with Saccharomyces boulardii CNCM I-745 and Lactiplantibacillus plantarum 299V. *Lwt* 2022, *157*, 113081. https://doi.org/10.1016/j.lwt.2022.113081.
- 105. Zhang, D.-N.; Guo, X.-Y.; Chen, Z.-G. A novel and efficient method for the isolation and purification of polysaccharides from lily bulbs by Saccharomyces cerevisiae fermentation. *Process Biochem.* **2014**, *49*, 2299–2304. https://doi.org/10.1016/j.procbio.2014.09.004.
- 106. Kuo-ChingWen; Lin, S.-P.; Yu, C.-P.; Chiang, H.-M. Comparison of Puerariae Radix and Its Hydrolysate on Stimulation of Hyaluronic Acid Production in NHEK Cells. *Am. J. Chin. Med.* **2010**, *38*, 143–155.
- 107. Yang, H.J.; Kwon, D.Y.; Moon, N.R.; Kim, M.J.; Kang, H.J.; Jung, D.Y.; Park, S. Soybean fermentation with Bacillus licheniformis increases insulin sensitizing and insulinotropic activity. *Food Funct.* **2013**, *4*, 1675. https://doi.org/10.1039/c3fo60198f.
- 108. Okamoto, T.; Sugimoto, S.; Noda, M.; Yokooji, T.; Danshiitsoodol, N.; Higashikawa, F.; Sugiyama, M. Interleukin-8 Release Inhibitors Generated by Fermentation of Artemisia princeps Pampanini Herb Extract With Lactobacillus plantarum SN13T. *Front. Microbiol.* **2020**, *11*, 1159. https://doi.org/10.3389/fmicb.2020.01159.
- 109. Lin, Y.-W.; Mou, Y.-C.; Su, C.-C.; Chiang, B.-H. Antihepatocarcinoma Activity of Lactic Acid Bacteria Fermented Panax notoginseng. *J. Agric. Food Chem.* **2010**, *58*, 8528–8534. https://doi.org/10.1021/jf101543k.
- 110. Jin, S.; Luo, M.; Wang, W.; Zhao, C.-j.; Gu, C.-b.; Li, C.-y.; Zu, Y.-g.; Fu, Y.-j.; Guan, Y. Biotransformation of polydatin to resveratrol in Polygonum cuspidatum roots by highly immobilized edible Aspergillus niger and Yeast. *Bioresour. Technol.* **2013**, *136*, 766–770. https://doi.org/10.1016/j.biortech.2013.03.027.
- 111. Sheih, I.C.; Fang, T.J.; Wu, T.-K.; Chang, C.-H.; Chen, R.-Y. Purification and Properties of a Novel Phenolic Antioxidant from Radix astragali Fermented by Aspergillus oryzae M29. *J. Agric. Food Chem* **2011**, *59*, 6520–6525. https://doi.org/10.1021/jf2011547.
- 112. Kwon, H.-K.; Jo, W.-R.; Park, H.-J. Immune-enhancing activity of *C. militaris* fermented with Pediococcus pentosaceus (GRC-ON89A) in CY-induced immunosuppressed model. *BMC Complement. Altern. Med.* **2018**, *18*, 75. https://doi.org/10.1186/s12906-018-2133-9.
- 113. Pyo, Y.; Kwon, K.H.; Jung, Y.J. Probiotic Functions in Fermented Foods: Anti-Viral, Immunomodulatory, and Anti-Cancer Benefits. *Foods* **2024**, *13*, 2386. https://doi.org/10.3390/foods13152386.
- 114. Li, K.; Park, M.-K.; Ngo, V.; Kwon, Y.-M.; Lee, Y.-T.; Yoo, S.; Cho, Y.-H.; Hong, S.-M.; Hwang, H.S.; Ko, E.-J.; et al. Lactobacillus plantarum DK119 as a Probiotic Confers Protection against Influenza Virus by Modulating Innate Immunity. *PLoS ONE* **2013**, *8*, e0075368. https://doi.org/10.1371/journal.pone.0075368.
- 115. Makino, S.; Sato, A.; Goto, A.; Nakamura, M.; Ogawa, M.; Chiba, Y.; Hemmi, J.; Kano, H.; Takeda, K.; Okumura, K.; et al. Enhanced natural killer cell activation by exopolysaccharides derived from yogurt fermented with Lactobacillus delbrueckii ssp. bulgaricus OLL1073R-1. *J. Dairy Sci.* 2016, 99, 915–923. https://doi.org/10.3168/jds.2015-10376.
- 116. Burrell, C.J.; Howard, C.R.; Murphy, F.A. Pathogenesis of Virus Infections. In *Fenner and White's Medical Virology*; Academic Press: Cambridge, MA, USA, 2017; pp. 77–104.
- 117. Zhao, W.; Wang, L.; Liu, M.; Zhang, D.; Andika, I.B.; Zhu, Y.; Sun, L. A Reduced Starch Level in Plants at Early Stages of Infection by Viruses Can Be Considered a Broad-Range Indicator of Virus Presence. *Viruses* **2022**, *14*, 1176. https://doi.org/10.3390/v14061176.
- 118. Zhu, S.; Viejo-Borbolla, A. Pathogenesis and virulence of herpes simplex virus. *Virulence* **2021**, *12*, 2670–2702. https://doi.org/10.1080/21505594.2021.1982373.

- 119. Michels, M.; Jesus, G.F.A.; Abatti, M.R.; Córneo, E.; Cucker, L.; de Medeiros Borges, H.; da Silva Matos, N.; Rocha, L.B.; Dias, R.; Simon, C.S.; et al. Effects of different probiotic strains B. lactis, L. rhamnosus and L. reuteri on brain-intestinal axis immunomodulation in an endotoxin-induced inflammation. *Mol. Neurobiol.* 2022, *59*, 5168–5178. https://doi.org/10.1007/s12035-022-02906-3.
- 120. Mousavi, E.; Makvandi, M.; Teimoori, A.; Ataei, A.; Ghafari, S.; Samarbaf-Zadeh, A. Antiviral effects of Lactobacillus crispatus against HSV-2 in mammalian cell lines. *J. Chin. Med. Assoc.* **2018**, *81*, 262–267. https://doi.org/10.1016/j.jcma.2017. 07.010.
- 121. Filip, R.; Gheorghita Puscaselu, R.; Anchidin-Norocel, L.; Dimian, M.; Savage, W.K. Global Challenges to Public Health Care Systems during the COVID-19 Pandemic: A Review of Pandemic Measures and Problems. *J. Pers. Med.* **2022**, *12*, 1295. https://doi.org/10.3390/jpm12081295.
- 122. Li, D.; Breiman, A.; le Pendu, J.; Uyttendaele, M. Anti-viral Effect of Bifidobacterium adolescentis against Noroviruses. *Front. Microbiol.* **2016**, *7*, 864. https://doi.org/10.3389/fmicb.2016.00864.
- 123. Choi, H.-J.; Song, J.-H.; Ahn, Y.-J.; Baek, S.-H.; Kwon, D.-H. Antiviral activities of cell-free supernatants of yogurts metabolites against some RNA viruses. *Eur. Food Res. Technol.* **2009**, *228*, 945–950. https://doi.org/10.1007/s00217-009-1009-0.
- 124. Santos, A.; San Mauro, M.; Sanchez, A.; Torres, J.M.; Marquina, D. The Antimicrobial Properties of Different Strains of Lactobacillus spp. Isolated from Kefir. Syst. Appl. Microbiol. 2003, 26, 434–437. https://doi.org/10.1078/072320203322497464.
- 125. Shojadoost, B.; Kulkarni, R.R.; Brisbin, J.T.; Quinteiro-Filho, W.; Alkie, T.N.; Sharif, S. Interactions between lactobacilli and chicken macrophages induce antiviral responses against avian influenza virus. *Res. Vet. Sci.* **2019**, *125*, 441–450. https://doi.org/10.1016/j.rvsc.2017.10.007.
- 126. Ermolenko, E.I.; Desheva, Y.A.; Kolobov, A.A.; Kotyleva, M.P.; Sychev, I.A.; Suvorov, A.N. Anti–Influenza Activity of Enterocin B In vitro and Protective Effect of Bacteriocinogenic Enterococcal Probiotic Strain on Influenza Infection in Mouse Model. *Probiotics Antimicrob. Proteins* 2018, 11, 705–712. https://doi.org/10.1007/s12602-018-9457-0.
- 127. Nalbantoglu, U.; Cakar, A.; Dogan, H.; Abaci, N.; Ustek, D.; Sayood, K.; Can, H. Metagenomic analysis of the microbial community in kefir grains. *Food Microbiol.* **2014**, *41*, 42–51. https://doi.org/10.1016/j.fm.2014.01.014.
- 128. Ang, L.Y.E.; Too, H.K.I.; Tan, E.L.; Chow, T.-K.V.; Shek, P.-C.L.; Tham, E.; Alonso, S. Antiviral activity of Lactobacillus reuteri Protectis against Coxsackievirus A and Enterovirus 71 infection in human skeletal muscle and colon cell lines. *Virol. J.* **2016**, *13*, 111. https://doi.org/10.1186/s12985-016-0567-6.
- 129. Kawashima, T.; Hayashi, K.; Kosaka, A.; Kawashima, M.; Igarashi, T.; Tsutsui, H.; Tsuji, N.M.; Nishimura, I.; Hayashi, T.; Obata, A. Lactobacillus plantarum strain YU from fermented foods activates Th1 and protective immune responses. *Int. Immunopharmacol.* 2011, 11, 2017–2024. https://doi.org/10.1016/j.intimp.2011.08.013.
- 130. Li, K.; Park, M.-K.; Ngo, V.; Kwon, Y.-M.; Lee, Y.-T.; Yoo, S.; Cho, Y.-H.; Hong, S.-M.; Hwang, H.S.; Ko, E.-J.; et al. Lactobacillus plantarum DK119 as a Probiotic Confers Protection against Influenza Virus by Modulating Innate Immunity. *PLoS ONE* **2013**, *8*, e75368. https://doi.org/10.1371/journal.pone.0075368.
- 131. Nakayama, Y.; Moriya, T.; Sakai, F.; Ikeda, N.; Shiozaki, T.; Hosoya, T.; Nakagawa, H.; Miyazaki, T. Oral administration of Lactobacillus gasseri SBT2055 is effective for preventing influenza in mice. *Sci. Rep.* **2014**, *4*, 4638. https://doi.org/10.1038/srep04638.
- 132. Botic, T.; Klingberg, T.; Weingartl, H.; Cencic, A. A novel eukaryotic cell culture model to study antiviral activity of potential probiotic bacteria. *Int. J. Food Microbiol.* **2007**, *115*, 227–234. https://doi.org/10.1016/j.ijfoodmicro.2006.10.044.
- 133. Ivec, M.; Botić, T.; Koren, S.; Jakobsen, M.; Weingartl, H.; Cencič, A. Interactions of macrophages with probiotic bacteria lead to increased antiviral response against vesicular stomatitis virus. *Antivir. Res.* **2007**, *75*, 266–274. https://doi.org/10.1016/j.antiviral.2007.03.013.
- 134. Grant, W.; Lahore, H.; McDonnell, S.; Baggerly, C.; French, C.; Aliano, J.; Bhattoa, H. Evidence that Vitamin D Supplementation Could Reduce Risk of Influenza and COVID-19 Infections and Deaths. *Nutrients* **2020**, *12*, 988. https://doi.org/10.3390/nu12040988.
- 135. Calder, P.; Carr, A.; Gombart, A.; Eggersdorfer, M. Optimal Nutritional Status for a Well-Functioning Immune System Is an Important Factor to Protect against Viral Infections. *Nutrients* **2020**, *12*, 1181. https://doi.org/10.3390/nu12041181.
- 136. Wu, G. Important roles of dietary taurine, creatine, carnosine, anserine and 4-hydroxyproline in human nutrition and health. *Amino Acids* **2020**, *52*, 329–360. https://doi.org/10.1007/s00726-020-02823-6.
- 137. Chen, Z.; Liang, W.; Liang, J.; Dou, J.; Guo, F.; Zhang, D.; Xu, Z.; Wang, T. Probiotics: Functional food ingredients with the potential to reduce hypertension. *Front. Cell. Infect. Microbiol.* **2023**, *13*, 1220877. https://doi.org/10.3389/fcimb.2023.1220877.
- 138. Voidarou, C.; Antoniadou, M.; Rozos, G.; Tzora, A.; Skoufos, I.; Varzakas, T.; Lagiou, A.; Bezirtzoglou, E. Fermentative Foods: Microbiology, Biochemistry, Potential Human Health Benefits and Public Health Issues. *Foods* **2020**, *10*, 69. https://doi.org/10.3390/foods10010069.